

Emerging Optical Methods for Endoscopic Barrett's Surveillance

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Abstract

Barrett's oesophagus is an acquired metaplastic condition that predisposes patients to the development of oesophageal adenocarcinoma, prompting the use of surveillance regimes to detect early malignancy for endoscopic therapy with curative intent. The currently accepted surveillance regime uses white light endoscopy together with random biopsies, but suffers poor sensitivity and discards information from numerous light-tissue interactions that could be exploited to probe structural, functional and molecular changes in the tissue. Advanced optical methods are now emerging that are exquisitely sensitive to these changes and hold significant potential to improve surveillance of Barrett's oesophagus if they can be applied endoscopically. The next decade will see some of these exciting new methods applied to Barrett's surveillance in new device architectures for the first time, potentially leading to a long-awaited improvement of the standard of care.

Introduction

Barrett's oesophagus is an acquired metaplastic condition that predisposes patients to the development of oesophageal adenocarcinoma (OAC)¹. The progression to cancer occurs through an intermediate stage known as dysplasia, which can be of low-grade (LGD) or high-grade (HGD). The cancer risk in non-dysplastic Barrett's oesophagus is estimated to range from 0.1 to 0.5%/year², but it increases to up to 9% in the presence of LGD³ and it is 4 times higher in patients harbouring HGD, compared to patients with LGD⁴. As the 5-year survival rate for oesophageal cancer is just 15%, but improves to 80% in patients with early-stage cancer^{5,6}, major advisory bodies recommend that patients with Barrett's oesophagus undergo routine endoscopic surveillance for signs of dysplasia or early carcinoma⁷⁻¹¹. Given the steep rise in cancer risk in patients with dysplasia, when LGD, HGD or in-situ carcinoma are detected there is indication to treat the early neoplastic lesion endoscopically with curative intent. Indeed, data from some retrospective studies indicate that endoscopic surveillance correlates with improved survival^{12,13}, although evidence from a case-controlled study¹⁴ did not confirm this and data from randomised controlled trials is lacking.

The current standard of care (SOC) for endoscopic surveillance uses high-definition white light endoscopy (HD-WLE) to identify suspicious lesions, then histopathological analysis of biopsied tissue for diagnosis (**Supplementary Figure 1**). To mitigate the risk of missing subtle, flat lesions, the protocol includes taking random biopsies at four-quadrant positions, in addition to targeted biopsies of visible lesions⁷. The resulting sensitivity is 40%-64% with specificity of 98-100%¹⁵, but the procedure is costly, time-consuming and prone to sampling error¹¹. The potential to improve clinical outcomes by detecting dysplasia with advanced optical methods has driven a great deal of research in this area.

White light endoscopy discards information from a wide range of contrast mechanisms (**Figure 1**) that can be exploited by more advanced optical methods to determine the disease state of tissue. In this review, we use the term 'optical method' to refer to the combination of an underlying contrast mechanism with an endoscope-compatible device, which results in a signal that can be used to guide the endoscopist. With demand for endoscopy predicted to rise substantially over the next decade¹⁶, the unmet clinical need for optical methods with improved diagnostic yield and lower device cost / complexity is particularly acute.

Advanced optical methods are categorised as 'red-flag', 'optical biopsy' or hybrid. Red-flag methods provide wide-field images and if they provide sufficient contrast for dysplasia, can replace the random four-quadrant biopsies through improved targeted biopsy. A recent study estimated that using a targeted biopsy protocol alone could reduce per-patient biopsy costs from ~£1000 to ~£30¹⁷. Conversely, optical biopsy methods measure a small area of tissue with the goal of providing *in vivo*, real-time diagnosis, which could ultimately replace physical biopsy and enable

surveillance and intervention to occur within the same procedure. Hybrid methods, as the name suggests, combine red flag and optical biopsy capabilities to identify and diagnose disease *in vivo*.

The field of advanced endoscopy has been described in several recent reviews^{9,15,18,19}. Here, we summarise the current status of the field with a focus on newly emerging optical methods, considering in particular the impact of device architecture on clinical translation. We conclude with a perspective on the potential for improvements in the endoscopic surveillance of Barrett's oesophagus.

Emerging endoscopic device architectures

At present, clinical endoscopic surveillance of Barrett's oesophagus is performed using a forward-facing trans-oral endoscope architecture, around which standard endoscopic tools, such as biopsy forceps and treatment devices, have been designed. Forward-facing endoscopes require articulation by the endoscopist to ensure complete surveillance of the tissue. A key consideration in the development and clinical translation of new optical methods is whether this remains the appropriate device architecture. An obvious and naïve approach to advanced endoscopy is simply to integrate the new optical imaging method into an existing forward-facing endoscope, to exploit familiarity of endoscopists with the presented images and retain access to the usual endoscopic tools. In recent years, research groups and commercial companies have taken a more 'out-of-the-box' approach, developing a host of alternative device architectures that overcome some or all of the limitations of forward-facing endoscopy, namely: low magnification; high procedure cost; the need for specialist operators; and restricted angular field of view (**Table 1 and Figure 2**).

Accessory channel endoscopes, or 'babyscopes', enable small-diameter probes to be inserted into standard forward-facing endoscopes through the channel that is normally used to introduce tools. They provide enhanced image data, often placed in direct contact with oesophageal tissue for optical biopsy methods. Unlike physical biopsy, increasing the number of these optical biopsies does not add significantly to procedure cost, however, the endoscopist must manually control the position the babyscope, so the results will still be subject to sampling error. In addition to this, the physiological movements of the oesophagus due to peristalsis and anatomic vicinity to the heart, make stabilization of the microscopic image challenging. In other cases, rather than direct contact, the imaging device employs a balloon that is inflated to ensure a fixed distance between the tissue and the central axis of the imaging hardware^{20,21}.

The high procedure cost of forward-facing endoscopy arises from the need for patient sedation in a specialist facility with a skilled endoscopist. Unsedated trans-nasal endoscopy (UTNE) provides standard endoscopy capabilities (imaging, articulation, insufflation, suction, biopsy) in a slim device that can be used without sedation, as the trans-nasal intubation does not involve contact with the root of the tongue and therefore does not trigger the

gagging reflex. UTNE has been successfully used for imaging BO and oesophageal dysplasia^{22–24}. Multiple UTNE systems are commercially available, including two disposable devices which make reprocessing feasible outside of a hospital environment²⁵. Recent studies using UTNE in screening for BO have found it to be comparable to standard endoscopy in clinical effectiveness, participation and safety²⁶ and considerably cheaper, especially if implemented in a mobile unit instead of a hospital²⁷. Nonetheless, UTNE image quality is currently insufficient for dysplasia detection in surveillance setting.

While UTNE still requires a skilled endoscopist, wireless capsule endoscopes are single-use, pill-shaped devices that can be administered by a non-specialist operator. Originally developed for small bowel imaging²⁸, wireless capsules have since been developed for the oesophagus²⁹. When surveyed, most patients prefer capsule endoscopy to regular endoscopy³⁰, which may improve adherence to surveillance protocols. Capsule endoscopy has become the gold standard for the small bowel, but studies in the oesophagus have yielded mixed results³¹. Wireless capsules have several significant limitations in the oesophagus³²: the need for a reclined ingestion protocol to increase the imaging period during swallowing from seconds to minutes³¹; difficulty in identifying the capsule location for a given image; and the inability to take biopsies during the procedure.

Tethered capsule endoscopes retain the benefits of wireless capsules while addressing several of their limitations in the oesophagus by using a cord to control the capsule's position³³. The tethered capsule is swallowed by the patient in an upright position, then imaging is performed while it is pulled back up from the stomach. The tethered architecture eliminates the risk of capsule retention and opens up the possibility of capsule re-use, which could lower per-procedure costs³⁴. A tethered capsule architecture for Barrett's surveillance is currently in clinical trials³⁵.

Finally, increased inspection time has been associated with an increased HGD/OAC detection rate in HD-WLE Barrett's surveillance³⁶, however, it is unclear whether this relates to longer time spent by the endoscopists in characterizing lesions detected during the examination. Inspection time is affected by the need of careful articulation of the endoscope to bring the entire luminal surface within the 160-180° forward-facing field of view. Wide angle endoscopes³⁷ with a 330° field of view have been successfully demonstrated in the colon. One study found that this decreased the colonic adenoma miss rate from 41% to 7%³⁸ compared to standard forward facing devices although this was not confirmed in a recent randomised study³⁹. Wide angle or stereoscopic devices⁴⁰, which allow 3D reconstruction, have yet to be implemented in the upper gastrointestinal tract but may improve surveillance.

Optical endoscopic methods used in clinical practice for BO surveillance

Several advanced optical methods, such as acetic acid chromoendoscopy, narrow band imaging and confocal laser endomicroscopy, have made their way into clinical use in some centres. Still, endoscopic practice varies significantly across countries and within the same country, and use of these advanced optical methods is often

restricted to tertiary referral centres delivering endoscopic treatment to a high volume of dysplastic patients. These methods have been extensively reviewed elsewhere^{19,41}, so we will only briefly summarise them here. For reference, **Table 2 and Supplementary Figure 2** show the current evidence, recommendation status, as well as key advantages and disadvantages of these methods. A recent meta-analysis by the American Society for Gastrointestinal Endoscopy (ASGE) suggest that to be recommended for targeting biopsy, a new technology should achieve at least 90% sensitivity, 80% specificity and 98% negative predictive value¹⁸.

Chromoendoscopy enhances contrast through topically applied dyes. Acetic acid eliminates the superficial mucosal layer and then causes acetylation of cellular proteins, resulting in whitening that highlights surface patterns. In case of neoplastic Barrett's, this is rapidly followed by focal erythema caused by vascular congestion in stromal capillaries, which is revealed as focal redness as loss of acetowhitening occurs¹⁷. These reactions are used to guide targeted biopsies and increase the yield of dysplasia, meeting the ASGE performance thresholds^{18,42}. Methylene blue chromoendoscopy has also been extensively investigated, but there are concerns regarding possible carcinogenic effects of the dye⁴³. Meta-analyses have found it to be inferior to WLE⁴⁴ and acetic acid chromoendoscopy¹⁸. It is therefore likely that acetic acid will become the standard conventional chromoendoscopic method for BO surveillance.

Virtual (also known as electronic or optical) chromoendoscopy improves contrast by modifying the endoscope hardware or software. This avoids the challenges of working with dyes, such as increased procedure time for dye administration and potential for adverse effects caused by the dye. Hardware modifications reported to date usually involve adapting the light source to focus on blue and green wavelength bands, where the haemoglobins are strongly absorbing, providing contrast based on changes in the tissue vasculature⁴¹. Narrow band imaging (NBI) is the most widely established and also meets the ASGE thresholds¹⁸. NBI highlights the capillary network of the superficial mucosa and the operator classifies the disease state of the tissue based on altered vascular and mucosal patterns associated with dysplasia⁴⁵. Blue laser imaging (BLI) is a similar technology that has also been tested in patients (*in vivo*, comparative study, n=39 patients)⁴⁶ and is under evaluation in Barrett's oesophagus⁴⁷. Software-based virtual chromoendoscopy methods^{48,49} use proprietary image processing algorithms to improve the contrast of mucosal and surface vessel patterns in the GI tract⁵⁰. While there is not currently sufficient data for advisory bodies to make recommendations¹⁸, clinical studies have shown that software-based approaches compare well to acetic acid chromoendoscopy (*in vivo*, prospective randomized pilot study, n=57)⁵¹. These early findings will need to be confirmed with large randomised controlled trials. Virtual chromoendoscopy has significant advantages in being label-free and easily applied in any WLE device architecture, including UTNEs²⁵ and capsules^{52,53}, so now has widespread availability.

While chromoendoscopy relies on light reflected from tissue, fluorescence imaging uses emission of a longer wavelength (or 'redder' colour) of light after illumination of the tissue to provide added contrast for dysplasia in endoscopic surveillance. Several structural and metabolic molecules intrinsic to tissue, such as collagen and NADH,

are fluorescent. Dysplastic tissue exhibits lower 'autofluorescence' than surrounding healthy tissue⁵⁴. Autofluorescence imaging (AFI) has high sensitivity for dysplasia, but low specificity because inflammation also reduces tissue autofluorescence⁵⁵. AFI is implemented by adding filters to the light source and detector on a standard endoscope, so has been combined with HD-WLE as well as virtual chromoendoscopy in endoscopic trimodal imaging (ETMI) in an effort to increase specificity. Trials to date have yielded mixed results^{56,57}; it remains unclear whether AFI truly adds to the already improved performance of NBI.

In addition to the intrinsic fluorescence, intravenous fluorescein (a fluorescent dye) can be used to highlight microvasculature and tissue structures to detect dysplasia. This is commonly examined using confocal laser endomicroscopy (CLE), which produces depth-sectioned, high magnification and resolution images, which can be used to spot changes in cell morphology associated with dysplasia, yielding high sensitivity and specificity¹⁸. An endoscope-based CLE system (eCLE) was recommended by the ASGE¹⁸, but requires a dedicated endoscope that is no longer on the market. A 'babyscope' probe-based CLE (pCLE) with lower resolution and limited depth sectioning, which can be inserted through the working channel of a standard forward facing endoscope is available. Clinical trial results to date indicate that pCLE can be used to identify neoplasia but is not yet sufficient to replace random biopsy⁵⁸. Neither fluorescence approach is available in UTE format as yet; the feasibility of incorporating fluorescence imaging into capsule endoscopes is being explored⁵⁹.

Emerging optical methods for endoscopic BO surveillance: what is on the horizon?

While existing advanced endoscopy methods have shown potential for improving the identification of dysplasia during BO surveillance, a number of exciting recent advances have been made in optical imaging that could address outstanding limitations in sensitivity and specificity, ultimately reducing the high miss rate^{15,60} (**Table 3 and Supplementary Figure 3**). Given the aforementioned challenges with the use of dyes and the other excellent recent reviews of optical molecular imaging⁶¹, we will concentrate here on label-free methods.

Interrogating disordered tissue structure

HD-WLE interrogates disordered tissue structure by presenting images of macroscopic abnormalities on the epithelial surface. Several recent advances have been made that allow endoscopists to probe cross-sectional information, up to several millimetres deep. Optical coherence tomography (OCT) can be thought of as 'optical ultrasound', with contrast derived from changes in refractive index rather than impedance mismatch. OCT uses scanning low-coherence interferometry to construct 3D reflectance images that reveal changes tissue microstructure arising due to variations in light scattering⁶², giving excellent contrast for dysplasia⁶³. Endoscopic applications of OCT

were made feasible by the shift from time-domain OCT to optical frequency domain imaging (OFDI), which significantly increased data acquisition rates. 3D images of the entire oesophagus can be acquired using an inflatable balloon babyscope device architecture, compatible with forward-facing endoscopes²⁰, or a rotating probe housed in a tethered capsule endoscope³³. Both helical, luminal imaging approaches can be referred to as volumetric laser endomicroscopy (VLE). VLE has been successfully correlated with histology in BO patients⁶⁴ (*ex vivo*, feasibility study, n=14 matched resection specimens) and detects oesophageal neoplasia *in vivo*⁶³ (*in vivo*, patient series, n=6 patients). One challenge with VLE is enabling guidance of tissue biopsy, which is not compatible with the existing device architectures; a combination of VLE and laser cautery has been shown to safely mark regions of interest for later biopsy under HD-WLE guidance⁶⁵ (*in vivo*, pilot study, n=22 patients). A second challenge remains with image interpretation; an experienced OCT endoscopist is currently needed, limiting widespread deployment. Automated image analysis is being investigated to alleviate this burden and bring the method closer to the clinic^{66,67}.

In addition to providing contrast for OCT, variation in light absorption and scattering from tissue can be recorded as a function of wavelength or angle. Diffuse reflectance spectroscopy (DRS), also called elastic scattering spectroscopy (ESS), illuminates the tissue with a standard white light source, but instead of collecting an image of the oesophagus using a camera, changes in the colour of the light arising from absorption and scattering events in the superficial layers of the tissue are measured with a spectrometer, a device that disperses white light into its component colours. Contact (ESS)⁶⁸ and fixed-distance (DRS)⁶⁹ babyscope probes can differentiate between healthy and dysplastic tissue in the oesophagus, though are typically restricted to point-based measurements rather than endoscopic imaging.

Taking the concept a step further, light scattering spectroscopy (LSS) singles out reflected light that has only scattered once in tissue. The benefit of this approach is that LSS measurements can be directly linked to tissue morphology via physical Mie scattering theory, enabling quantitative measurements of the size and density of cell nuclei, which is associated with disease state⁷⁰. An early LSS study achieved 90% sensitivity and specificity for oesophageal dysplasia⁷¹ (*in vivo*, single centre pilot study, n=13 patients, n=76 sample positions) with a babyscope contact probe, but unwanted variations in probe-tissue separation led to challenges for interpretation. Hardware developments overcame this limitations to enable 8 cm segments of oesophagus to be mapped with 92% sensitivity and 96% specificity⁷² (*in vivo*, single centre pilot study, n=9 patients, n=95 biopsies), showing potential for this to become a useful red-flag tool for guiding targeted biopsies in Barrett's surveillance. Angle-resolved low coherence interferometry (a/LCI) also looks at singly-scattered light but probes the angular scattering distribution of just a single colour of light. a/LCI has been shown to identify dysplasia (including LGD) with 100% sensitivity and 84% specificity *in vivo*⁷³ (*in vivo*, 2 centre pilot study, n=46 patients, n=172 sample positions) and a negative predictive value of 100%.

Although DRS, LSS and a/LCI were originally point measurement methods, the ability to provide 2D maps that co-register with HD-WLE or other images of tissue anatomy has now been demonstrated^{69,74}, although not yet tested

in vivo. There are also phase and polarisation-sensitive endoscopic methods on the horizon that derive contrast from scattering and present wide-field images^{75,76}. While VLE is the most advanced method for interrogating microstructure in terms of clinical translation, if their performance remains high in randomised controlled trials, DRS, LSS and a/LCI have potential to become valuable tools to guide targeted biopsy.

Interrogating abnormal tissue function and metabolism

Virtual chromoendoscopy has been successful in improving targeted biopsy based on changes in tissue vasculature, but only interrogates superficial epithelial changes. OCT has shown exciting possibilities for cross-sectional imaging of tissue function, using measurements of blood flow to highlight the vasculature^{77,78}, although these have yet to be clinically validated. Another method that provides cross-sectional vascular information is photoacoustic endoscopy (PAE), which uses optical excitation of tissue to generate ultrasound⁷⁹. The benefit of this approach is that highly optically absorbing molecules in tissue, such as haemoglobins, can be resolved at far greater penetration depth than is available from exclusively optical imaging. PAE could therefore directly provide high resolution cross-sectional virtual chromoendoscopy at centimetre depths, allowing visualisation of vascular patterns associated with dysplasia. PAE devices using a similar helical scanning implementation to OCT^{80–82} have been applied in rabbit oesophagi⁸³, but further development is needed to increase radial resolution and acquisition speed, as well as to address challenges with image interpretation (similar to those of OCT). Recent successful studies using photoacoustic imaging in breast cancer diagnosis and other areas⁸⁴ suggests that application of PAE in Barrett's surveillance may yet yield valuable information, potentially in combination with OCT and other advanced methods⁸¹.

Using a contrast mechanism already applied in the clinic, time-resolved assessment of tissue autofluorescence may overcome the present challenge of poor specificity in the interpretation of AFI. Changes in autofluorescence intensity, as measured by standard AFI endoscopes, can be confounded by surface irregularities and non-uniform illumination. Fortunately, measuring the lifetime of the fluorescence signal, rather than its absolute intensity, avoids these confounding factors⁸⁵. Fluorescence lifetime microscopy (FLIM) is able to map changes in local tissue microenvironment and has shown promise in detection of cancers in *ex vivo* and *in vivo* studies⁸⁵. For many years, the clinical translation of FLIM was limited by the size, cost and complexity of the instrumentation and the need for long integration times due to weak signals. A 2003 study of point-based fluorescence lifetime measurements found sensitivity and specificity for HGD of less than 60% using time resolved fluorescence⁸⁶ (*in vivo*, single centre pilot study, n=37 patients, n=108 fluorescence decay profiles). More recently, however, compact diode-pumped laser-based excitation sources and time-gated methods have addressed instrumentation limitations⁸⁷ meaning wide-field FLIM endoscopes with near-video rate acquisition ($\sim 2\text{Hz}$)^{88–90} are now available. While these have been used *in vivo*^{88,91} they have yet to be applied to Barrett's surveillance and may yield improved performance in this context.

Complementing wide-field FLIM approaches, multi-photon microscopy (MPM) provides an autofluorescence-based alternative to pCLE. MPM is a scanning, optical sectioning, imaging approach in which fluorescence is spatially delineated using non-linear optical excitation. *Ex vivo* MPM of fresh punch biopsies can successfully distinguish squamous mucosa, gastric columnar mucosa and intestinal metaplasia⁹² (*ex vivo*, n=25 patients, n=35 biopsies) suggesting that MPM could be used to identify dysplasia. MPM endoscopes are being developed and can incorporate additional features from microscopy such as super-resolution imaging⁹³. Although MPM is at a very early stage of development, it holds potential to perform high magnification, depth-sectioned, label-free endomicroscopy as part of Barrett's surveillance.

Interrogating bulk molecular composition

Changes in bulk molecular composition can be determined using the spectral 'fingerprint' measured through Endoscopic Raman Spectroscopy (ERS), which is typically classified based on machine-learning methods that use a training set of spectra where the disease classification is known from histopathology analysis⁹⁴. ERS is sensitive to the abundance of molecular bonds, primarily lipid, protein and nucleic acid content in tissue. ERS probes are typically introduced through a babyscope into a standard forward-facing endoscope and positioned directly on a suspicious region of tissue to provide a point-based measurement.

The low intensity of Raman signals has been a hurdle for ERS, historically resulting in very slow data acquisition. Nonetheless, Bergholt et al. recently demonstrated an ERS babyscope system, including a classification algorithm based on a Raman library of >12000 spectra, that could differentiate between columnar lined epithelium, non-dysplastic BO or HGD, in real time (0.2 sec), passing this information to the endoscopist using auditory feedback⁹⁵ (*in vivo*, pilot study, n=77 patients, sensitivity 87.0%, specificity 84.7%). Trans-nasal image-guided Raman spectroscopy has also been demonstrated⁹⁶. Alternatively, Coherent Anti-stokes Raman Spectroscopy (CARS), which uses multiple photons to probe specific regions of the spectral fingerprint, has been suggested as a way to overcome the low Raman intensity and several prototype endoscopes have been developed, despite challenges associated with non-linear effects in fibres and the design of miniature optics^{97,98}. Precision remains a challenge for ERS and ECARS, due to pressure-based signal variation, but prospective randomized multicentre trials are underway⁹⁵ and further devices are under development⁹⁹. Raman spectroscopy is thus a promising method that could provide point measurements for optical biopsy and be scanned to assess larger areas of tissue if biopsy guidance were desired.

Multimodal methods

The recent advances highlighted above suggest that the intrinsic optical interactions with tissue have the potential to improve the diagnostic yield of Barrett's surveillance. Naturally, combining several of these into a single device architecture could have added benefits, giving access to structural, functional and molecular information simultaneously. For example, a recent pilot study with an intraoperative fibre probe combining DRS, ERS and fluorescence spectroscopy achieved 100% sensitivity and 93% specificity for several cancers¹⁰⁰.

Achieving a successful combination, however, requires careful optical design and often complex instrumentation. One promising route to overcoming this challenge may lie in the use of hyperspectral imaging (HSI), where the light illuminating the tissue and being imaged is dispersed into its component colours, or rapidly modulated. NBI is a simple example of this, where restricting illumination to two wavelengths highlights the vasculature in tissue due to strong haemoglobin absorption. HSI goes further, recording 10s or 100s of colours at every pixel in an endoscopic image, which can then be processed using machine-learning methods to resolve reflectance (e.g. NBI, BLI, DRS), fluorescence (AFI) or Raman (ERS) information into separate images. HSI has shown potential for aiding cancer diagnosis in a range of organ sites, including the oesophagus¹⁰¹, although has yet to be demonstrated through *in vivo* trials in Barrett's surveillance. HSI hardware is often bulky and slow; optimisation of the HSI hardware, for example using compact spectrally resolved detector arrays¹⁰², may assist in the future with real-time clinical application.

Introducing cross-sectional methods into such endoscopes adds a further challenge, particularly if helical scanning implementations are needed, as these typically require further endoscopist training to develop specialist expertise for interpretation. If true optical biopsy is to be achieved, cross-sectional information will be vital. The addition of scattering measurements that exploit other dimensions of light than colour, such as phase and polarisation^{75,76} may help to achieve accurate depth-sectioning and hence improved identification of early dysplasia in the oesophagus.

Translational outlook for new optical methods

Novel optical imaging methods that probe structural, functional and molecular information in tissue hold significant potential to improve BO surveillance endoscopy. Clinical translation of these emerging optical methods, however, faces many challenges. The most appropriate device architecture for implementation of the optical method must first be identified. Next, the resulting endoscopic instrument must undergo technical and biological validation in phantoms, preclinical models and clinical trials to ensure performance and safety standards are met¹⁰³. Several challenges can be encountered at this stage. For technical validation, the lack of calibration standards and accepted

internal exposure limits for optical diagnostic methods makes it difficult to assess the risks of potential thermal damage or photoallergic reactions for a new device¹⁰⁴. Testing in *ex vivo* tissue does not provide an appropriate reference in this regard, since blood flow will dissipate heat. Furthermore, for biological validation, the optical properties of tissue can change markedly when examined *ex vivo*, which may limit the utility of subsequent *in vivo* findings^{92,105}. Given the limited number of studies detailing the nature of changes in light-tissue interactions during the development of dysplasia, particular benefit for biological validation may be derived from further *ex vivo* and *in vivo* analysis of oesophageal tissue in different disease states.

Clinical trials at expert research centres with enriched populations or complex protocols may also bias findings, which can lead to disappointing results once deployment is more widespread^{56,57}. For example, ASGE recommendations for use of acetic acid chromoendoscopy, NBI and eCLE for targeted biopsy assume the endoscopist has specialist training in image acquisition and interpretation for these methods. The use of histopathological diagnosis as the gold standard for evaluation of a new optical endoscopic device can also be confounding, since it itself is prone to sampling and interpretation errors^{11,106}. Ideally, biopsies would be taken under guidance of a new optical method and subjected to consensus histopathology to minimise such errors. The alternative is to establish longitudinal studies that relate early-stage imaging parameters to late-stage clinical outcomes, which is extremely costly. Once devices and specialists are available across multiple sites, metrics to interpret the images must be developed and perfected, often by consensus of an expert group. This complex development process requires strong multidisciplinary collaboration, including expertise from medicine, engineering, physics, biomedical sciences, computer science and mathematics. Clear and open communication between those developing new endoscopic devices and those who will ultimately operate them is paramount.

The emerging optical methods reviewed here aim to either increase the contrast of wide-field ‘red flag’ endoscopic surveillance, for improved targeting of physical biopsies, or to provide an ‘optical biopsy’ that could yield diagnostic information directly during the endoscopic procedure. Enhancing the existing red-flag forward-facing endoscopy could be achieved through the addition of light scattering spectroscopy⁷², but further clinical studies are needed to establish potential for improved BO surveillance. Hyperspectral endoscopy has the potential to enable a truly multi-modal red-flag and is the subject of ongoing work across a number of centres.

The majority of optical methods reviewed provide an optical biopsy. Several spectroscopic methods (DRS, LSS, a/LCI, ERS, ECARS) are compatible with forward-facing endoscopy using a babyscope and have the potential to be automated to give a fast real-time binary feedback to the endoscopist^{73,95}. As their application becomes more widespread, the acquisition of more data will enable refinement of the automated classification algorithms, further improving sensitivity and specificity. Imaging methods applied for optical biopsy, such as CLE and MPM, also hold promise, particularly if they are able to achieve adequate depth sectioning to yield high quality images for

interpretation by histopathologists. Given the restricted field-of-view of such methods, they remain reliant on a high contrast red-flag endoscopy to achieve their diagnostic potential.

Hybrid approaches that combine red-flag and optical biopsy information, as well as structural and functional information, have also been demonstrated. Photoacoustic endoscopy is at an early stage, while volumetric laser endomicroscopy (VLE) is the most mature of the emerging methods described in this review, with a commercial system already available²⁰. Furthermore, it has been demonstrated in both babyscope balloon and tethered capsule device architectures. If contact between the capsule device and the lumen can be maintained, VLE is able to capture high-resolution volumetric images of the entire oesophagus, an interesting niche. Although interpretation is currently performed offline, the development of automated diagnosis algorithms⁶⁶, accelerated by recent advances in machine learning coupled with increasingly inexpensive computing power, offers the possibility of a real-time computer-aided diagnosis. Such information could be combined with immediate laser cautery marking of suspicious lesions⁶⁵. Though at present a standard forward-facing endoscope would still be required for physical biopsy and therapeutic intervention, the comprehensiveness, simplicity, and apparent achievability of VLE is exciting.

In summary, an ideal BO endoscopic surveillance method would perform comprehensive investigation of the oesophagus with high sensitivity and specificity for dysplasia. It should allow for use of endoscopic tools for marking and biopsy if necessary. It should also be possible to implement with minimal additional training of endoscopist operators and image interpreters. To achieve widespread deployment in healthcare systems, no significant change to procedure times or costs should be made; ideally these would be reduced. If possible, availability of endoscopy to BO patients should be increased, for example by enabling deployment by non-specialist operators in primary care centres, and physical discomfort with the procedure should be decreased, to improve compliance with surveillance programmes.

Though several advanced endoscopy methods have been recommended for routine use in Barrett's oesophagus, histological assessment of HD-WLE targeted and random biopsies is still the standard-of-care. Many emerging methods combine a contrast mechanism and device architecture to achieve some of the aforementioned requirements, but fall short of providing the ideal solution. Continuing advances in hardware and software are allowing endoscopic application of optical methods developed for other indications. The next decade will see some of these exciting new methods applied to Barrett's surveillance in new device architectures for the first time, potentially leading to a long-awaited improvement of the standard of care.

Acronyms

4QB	4-quadrant biopsies
a/LCI	angle-resolved low coherence interferometry
AA	Acetic Acid
ACG	American College of Gastroenterologists ⁹
AFI	autofluorescence imaging
AGA	American Gastroenterology Association ¹⁰⁷
ASGE	American Society for Gastrointestinal Endoscopy ¹⁸
BO	Barrett's Oesophagus
BSG	British Society of Gastroenterologists ⁷
CARS	coherent anti-Stokes Raman spectroscopy
DRS	diffuse reflectance spectroscopy
eCLE	endoscope-based confocal laser endomicroscopy
ERS	endoscopic Raman spectroscopy
ESGE	European Society of Gastrointestinal Endoscopy ⁸
ESS	elastic scattering spectroscopy
ETMI	endoscopic trimodal imaging

FICE	Fujicon Intelligent Colour Enhancement
FLIM	fluorescence lifetime imaging
HD-WLE	high-definition white light endoscopy
LSS	light scattering spectroscopy
MB	methylene blue
MPM	multi-photon microscopy
MSI/HIS	multi-/hyper- spectral imaging
NBI	narrow band imaging
OCT	optical coherence tomography
OFDI	optical frequency domain imaging
OMI	optical molecular imaging
PAE	photoacoustic endoscopy
pCLE	probe-based confocal laser endomicroscopy
SOC	standard of care
TRF	time resolved fluorescence
VLE	volumetric laser endomicroscopy

Figures

Figure 1. Contrast mechanisms. An optical contrast mechanism consists of three elements: illumination, interaction, and detection of light. By carefully controlling the properties of the light illuminating the tissue (left) we can probe specific light-tissue interactions (right). These include reflection, absorption, elastic/inelastic scattering and fluorescence. Advanced endoscopic imaging modalities use these interactions as a source of contrast for detection of dysplasia (centre). Information about the interaction is encoded within the properties of the output light: the wavelength, the distance between peaks in the wave; the polarisation, the direction in which the wave oscillates; the phase, the point in the cycle of the wave; and the intensity, the power within the wave. Detection of these properties allows us to infer information about tissue disease state.

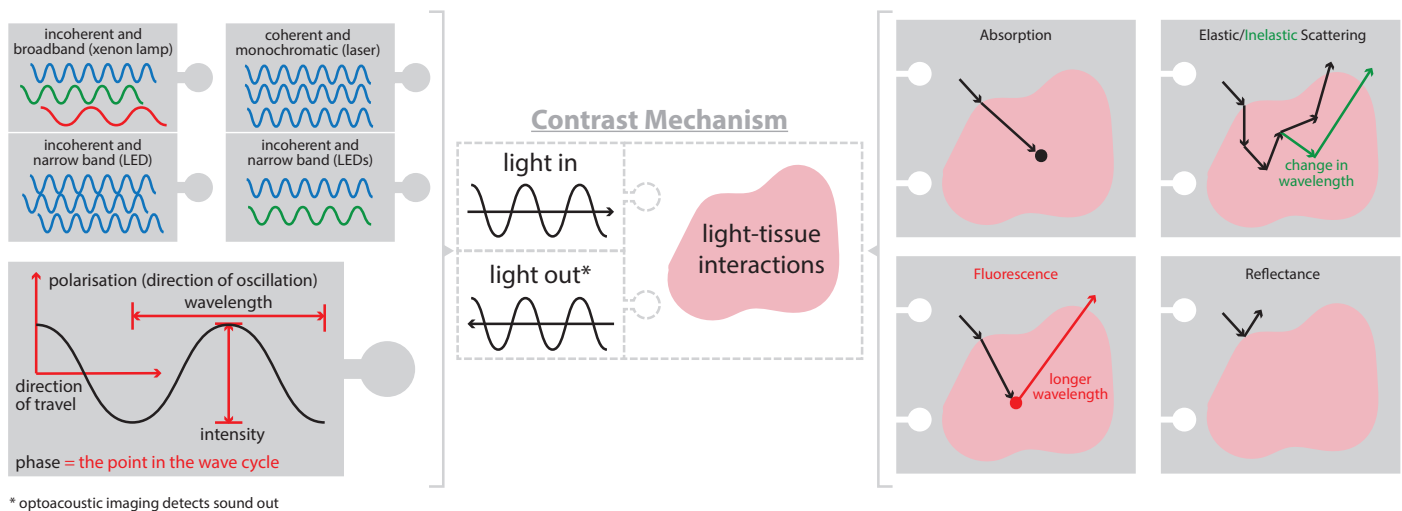


Table 1. Endoscopic device architectures.

** Theoretically most combinations of contrast mechanism and device type are possible. Here we give the contrast mechanisms that are most compatible with the advantages and disadvantages of the device architecture.

*** Image type is again dependent on contrast mechanism. Here we give the image types for the most compatible contrast mechanisms for the device architecture.

Schematics of each device architecture are shown in Figure 2.

Device Architecture	Example Device (s)	FOV	Advantages/Disadvantages	Most Compatible Contrast Mechanism**	Typical Image Type***	Ref
Forward Facing (Trans-oral)	Standard commercial endoscopes e.g. Olympus, Pentax, Fuji	Wide (typically 140° luminal view)	<ul style="list-style-type: none"> + Availability + Familiarity + Wide variety of tools for biopsy, washing, marking + Articulation – Endoscopist must articulate to survey entire surface 	WLE, Chromoendoscopy, NBI, eCLE, OMI, MSI/HSI, Polarimetry	En-face, luminal	
Forward Facing (Trans-nasal)	Standard commercial endoscopes e.g. Olympus, Pentax, Fuji	Wide (typically 140° luminal view)	<ul style="list-style-type: none"> + Improved patient tolerance and no sedation required + Articulation + Shorter, less costly procedure – Endoscopist must articulate to survey entire surface – Lower quality image*, narrower working channel inappropriate for interventions, poorer suction and air function and smaller biopsy capabilities compared with trans-nasal endoscopes *unsuitable for Barrett's surveillance 	WLE, NBI	En-face, luminal	108
Babyscope E.g. Contact Probe	Mauna Kea Cellvizio®	Narrow (10s – 100s microns)	<ul style="list-style-type: none"> + Compatible with insertion through working channel of standard endoscopes – Must be used alongside standard endoscope for articulation, washing, biopsy, marking – Contact with lumen must be carefully controlled – Small FOV 	pCLE, ERS, ESS/DRS, a/LCI, MSI/HSI, FLIM, MPM, PA, Polarimetry	Spectrum, en-face	109
Balloon Based	NinePoint NvisionVLE®	Volumetric	<ul style="list-style-type: none"> + Controlled withdrawal + Potential for cautery marking capability + Compatible with insertion through working channel of standard endoscopes + Allows full volumetric imaging of oesophagus – No biopsy, washing capabilities – Contact with lumen must be carefully controlled 	OCT/VLE/OFDI	Volumetric	20,21
Wireless Capsule	Given Imaging PillCam® ESO series	2 x 169° (ESO2)	<ul style="list-style-type: none"> + No sedation required + Can be implemented in primary care + Potential for low cost if reusable – One shot (cannot return to suspicious lesions) – No biopsy, washing, marking capabilities – Long delay for capsule to pass (8 – 10 hours) – No control over motion – Contact with lumen must be carefully controlled 	WLE, NBI, MSI/HSI, Polarimetry	En-face, luminal or circumferential	29,30,110
Tethered Capsule	No commercial devices	Volumetric	<ul style="list-style-type: none"> + No sedation required + Can be implemented in primary care + Potential for low cost if reusable + Controlled withdrawal + Potential for cautery marking capability + Immediate removal of capsule + Allows full volumetric imaging of oesophagus – No biopsy, washing capabilities – Contact with lumen must be carefully controlled 	OCT/VLE/OFDI	Volumetric	35
Wide Angle	EndoChoiceFuse (330°)	Extra Wide (>140°)	<ul style="list-style-type: none"> + Familiarity + Wide variety of tools for biopsy, washing, marking + Articulation + Wide FOV allows viewing of entire lumen with minimal articulation – Increased cost 	WLE, Chromoendoscopy, NBI, eCLE, OMI, MSI/HSI, Polarimetry	En-face, Circumferential	37

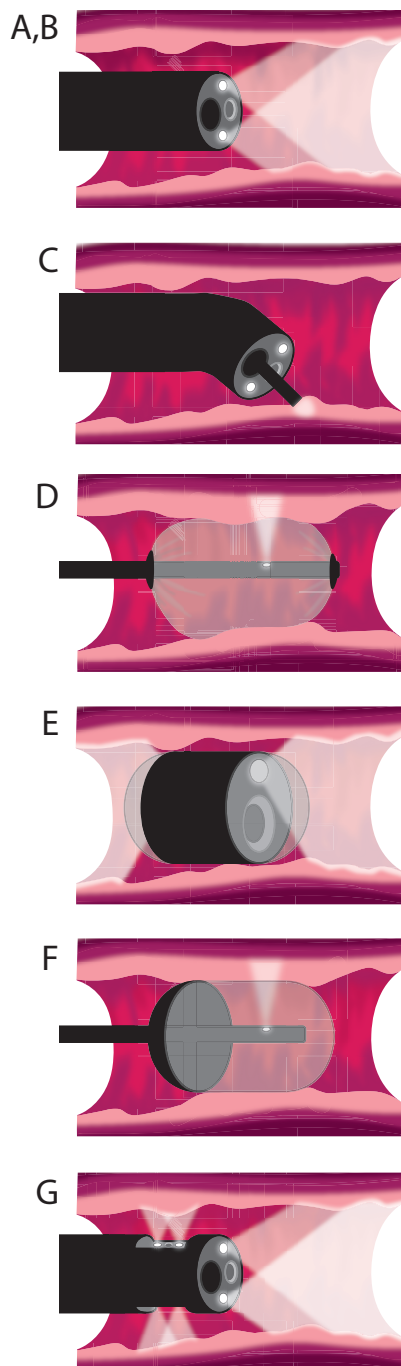


Figure 2. Schematic representations of the endoscopic device architectures listed in Table 1.

- A: Forward Facing (Trans-oral)**
- B: Forward Facing (Trans-nasal)**
- C: Babyscope E.g. Contact Probe**
- D: Balloon Based**
- E: Wireless Capsule**
- F: Tethered Capsule**
- G: Wide Angle**

Table 2. Advanced optical methods in clinical use for endoscopic BO surveillance.

Example images for each method are shown in **Supplementary Figure 2**.

* cannot be advocated or discouraged at this time

	Advantages	Disadvantages	Sensitivity	Specificity	Recommendation				
					BSG	ASG	ACG	AGA	ESG
HD-WLE + 4QB targeted biopsies and histopathology	<ul style="list-style-type: none"> Widely available Well established 	<ul style="list-style-type: none"> Prone to sampling error¹¹ Exhaustive biopsies are expensive 	0.40-0.68 ¹⁵	0.98-1.00 ¹⁵	✓	✓	✓	✓	✓
Chromoendoscopy	<ul style="list-style-type: none"> Inexpensive Widely available AA has shown high sensitivity and specificity for detecting dysplasia¹⁸ 	<ul style="list-style-type: none"> Potential toxicology issues⁴³ (MB) Increase in procedure time¹⁸ Low inter-observer agreement No current procedural terminology for billing and reimbursement¹⁸ Difficulty in achieving uniform application of dye¹¹ 	0.92-0.966 (AA) 0.642 (MB) ^{18,111}	0.846-0.96 (AA) 0.959 (MB) ^{18,111}	X	✓ (AA)	X	X	X
Hardware-based Virtual Chromoendoscopy (e.g. NBI, BLI)	<ul style="list-style-type: none"> Ability to visualise mucosal and vascular patterns Widely available Ease of use High sensitivity and specificity for detecting HGD^{112,113} Reduced number of biopsies¹¹⁴ 	<ul style="list-style-type: none"> No universal classification criteria until recent BING criteria⁴⁵ Low inter-observer agreement Low sensitivity for LGD¹¹⁵ 	0.942 ¹⁸	0.975 ¹⁸	X	✓	X	*	X
Software-based Virtual Chromoendoscopy (e.g. FICE, iSCAN)	<ul style="list-style-type: none"> No additional hardware costs 	<ul style="list-style-type: none"> Lack of data 	0.83 ⁵¹ (HGD, FICE)	Unavailable	X	X	X	*	X
Autofluorescence Imaging (AFI)	<ul style="list-style-type: none"> Easy to combine with NBI and WLE 	<ul style="list-style-type: none"> Many studies biased by comparison with substandard WLE¹¹⁶ Limited value in routine surveillance¹¹⁶ 	0.50 ¹¹⁷ (HGD)	0.61 ¹¹⁷ (HGD)	X	X	X	X	X
Endoscopic Trimodal Imaging (ETMI)	<ul style="list-style-type: none"> Reduced false positive rate relative to AFI alone 	<ul style="list-style-type: none"> Useful in tertiary referral centres⁵⁶ but not in community practice⁵⁷ 	0.805 ¹⁸	0.46 ¹⁸	X	X	X	X	X
Probe-based Confocal Laser Endomicroscopy (pCLE)	<ul style="list-style-type: none"> Probe can be inserted through working channel of standard endoscope Close to in-vivo histology 	<ul style="list-style-type: none"> Often uses exogenous contrast (fluorescein) 	0.903 ¹⁸	0.773 ¹⁸	X	X	X	X	X

**Endoscope-based
Confocal Laser
Endomicroscopy
(eCLE)**

- Close to in-vivo histology

- Requires dedicated endoscope (in contrast to pCLE)
- Often uses exogenous contrast (fluorescein)

0.904¹⁸

0.927¹⁸

x ✓ x x x

Table 3. Emerging optical methods for endoscopic BO surveillance.

Example images for each method are shown in **Supplementary Figure 3**.

Blue: *Exogenous contrast*

Red: *Interrogating disordered tissue microstructure*

Green: *Interrogating abnormal tissue function and metabolism*

Purple: *Interrogating bulk molecular composition*

Orange: *Multimodal methods*

	Source of Contrast	Biological Change in Cancer	Functional Information	Morphological Information	Depth Sectioning	Strengths/Weaknesses	Status/Prospect
Optical Molecular Imaging (OMI)	Exogenous fluorophores conjugated to targeting moieties (lectins, peptides, antibodies, affibodies, enzymes) that targeting intracellular and extracellular proteins and enzymes	Biochemical	✓	✓	✗	+ Specificity – Exogenous contrast – Surface images – Cost	<i>In vivo</i> trials in BO ⁶¹ . Potential to be translated for wide field surveillance. Awaiting further <i>in vivo</i> trials.
Optical Coherence Tomography/ Optical Frequency Domain Imaging/ Volumetric Laser Endomicroscopy (OCT/OFDI/VLE)	Structural features.	Organizational changes, vasculature	✗	✓	✓	+ High resolution + Depth sectioning + Endogenous contrast – Large image datasets	<i>In vivo</i> trials in BO (patient series, n=6) ⁶³ .
ElastiC Scattering Spectroscopy/ Diffuse Reflectance Spectroscopy/ Light Scattering Spectroscopy (ESS/DRS/LSS)	Endogenous scatterers (cells, nuclei, organelles)	Biochemical, cellular/organelle changes	✓	✗	✗	+ Depth penetration + Endogenous contrast – Spectrum rather than image	<i>In vivo</i> trials in BO (single centre pilot study, n=9 patients, n=95 biopsies) ⁷² . No trials published in last 10 years.
Angle-resolved Low Coherence Interferometry (a/LCI)	Nuclei	Increase in nuclear size	✗	✓	✗	+ High sensitivity and specificity in pilot study + Endogenous contrast – Tissue orientation can affect results	<i>In vivo</i> pilot study in BO (2 centre pilot study, n=46 patients, n=172 sites) ⁷³ . Combination with OCT. Clinical trials likely.
Polarimetry	Nanostructural anisotropy	Collagen, cellular orientations, organizational changes	✓	✗	✗	+ Endogenous contrast + Instrumentation challenges	No trials in BO. Awaiting further device development.

Photoacoustic Endoscopy (PAE)	Endogenous absorbers (NAD(P)H, haemoglobin)	Vasculature	✓	✓	✓	<ul style="list-style-type: none"> + Volumetric images + Endogenous contrast – Instrumentation challenges – Limited resolution at present – Long acquisition times 	<i>In vivo</i> imaging of oesophagus in animals ⁸³ . No trials in BO. Awaiting application to BO.
Fluorescence Lifetime Imaging (FLIM)	Endogenous fluorophores (NAD(P)H, flavins, collagen, elastin, phenylalanine, tryptophan, tyrosine, melanin).	Biochemical, microenvironment (pH, [O ₂], [Ca ²⁺])	✓	✓	✓	<ul style="list-style-type: none"> + More robust than traditional AFI + Endogenous contrast – Safety of UV illumination – Long acquisition times 	<i>Ex vivo</i> trials (TRF) (single centre pilot study, n=37 patients, n=108 fluorescence decay profiles) ⁸⁶ . Awaiting in vivo trials.
Multi-photon Microscopy (MPM)	Endogenous fluorophores (NAD(P)H, flavins, collagen, elastin, phenylalanine, tryptophan, tyrosine, melanin).	Cell type	✓	✓	✓	<ul style="list-style-type: none"> + Depth sectioning + High resolution + Endogenous contrast – Requires high illumination intensity 	<i>Ex vivo</i> trials in BO (n=25 patients, n=35 biopsies) ⁹² . Awaiting in vivo trials.
Endoscopic Raman Spectroscopy (ERS)	Specific molecular groups (e.g. C-C proteins, C-C ring of phenylalanine, C-C of lipids, C-N and N-H of proteins, CH ₂ of lipids, C=C of porphyrins, C=O of proteins and lipids)	Biochemical	✓	✗	✗	<ul style="list-style-type: none"> + Detailed biochemical information + Algorithms have been developed + Multicentre trials underway – Endogenous contrast. – Spectrum rather than image – Repeatability has been questioned – Validation using ex-vivo tissue is difficult 	<i>In vivo</i> trials in BO (pilot study, n=450 patients) ⁹⁵ . Potential to be translated if repeatability can be confirmed. Awaiting multicentre trials.
Coherent anti-Stokes Raman Spectroscopy (CARS)	Specific molecular groups (see ERS)	Biochemical	✓	✓	✓	<ul style="list-style-type: none"> + Detailed biochemical information + Increased sensitivity compared to ERS – Instrumentation challenges – Requires high illumination intensity 	No trials in BO. Awaiting further device development.
Multi-/Hyper- Spectral Imaging (MSI/HIS)	Endogenous chromophores (NAD(P)H, haemoglobin)	Vasculature, visible lesions	✓	✓	✗	<ul style="list-style-type: none"> + Simple + Compact + Endogenous contrast – Surface images 	No trials in BO. Awaiting application to BO.

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Contributors

DJW, CRMF and SEB wrote the manuscript, with feedback and clinical guidance from MdP.

Declaration of interests

SEB receives research support from iThera Medical GmbH and PreXion Inc. for photo acoustic imaging studies beyond the scope discussed in this review. The other authors declared no conflicts of interest.

Search strategy and selection criteria

We searched PubMed, Scopus and Google for articles published up to Nov 1, 2017 using the terms, “imaging”, “Barrett’s”, “endoscope”, “capsule”, “optical” and “detection”. Additional articles were also identified through searches of the references of these articles. Only papers in English were reviewed. The final reference list was generated on the basis of originality, impact and relevance to the aims of this review.

References

- 1 Gatenby P, Caygill C, Wall C, Bhattacharjee S, Ramus J, Watson A *et al.* Lifetime risk of esophageal adenocarcinoma in patients with Barrett's esophagus. *World J Gastroenterol* 2014; **20**: 9611–7.
- 2 Desai TK, Krishnan K, Samala N, Singh J, Cluley J, Perla S *et al.* The incidence of oesophageal adenocarcinoma in non-dysplastic Barrett's oesophagus: a meta-analysis. *Gut* 2012; **61**: 970–6.
- 3 Duits LC, Phoa KN, Curvers WL, Ten Kate FJW, Meijer G a, Seldenrijk C a *et al.* Barrett's oesophagus patients with low-grade dysplasia can be accurately risk-stratified after histological review by an expert pathology panel. *Gut* 2015; **64**: 700–6.
- 4 Reid BJ, Levine DS, Longton G, Blount PL, Rabinovitch PS. Predictors of progression to cancer in Barrett's esophagus: Baseline histology and flow cytometry identify low- and high-risk patient subsets. *Am J Gastroenterol* 2000; **95**: 1669–1676.
- 5 CRUK. Oesophageal cancer statistics. <http://www.cancerresearchuk.org/health-professional/cancer-statistics/statistics-by-cancer-type/oesophageal-cancer>. .
- 6 Barbour AP, Jones M, Brown I, Gotley DC, Martin I, Thomas J *et al.* Risk stratification for early esophageal adenocarcinoma: analysis of lymphatic spread and prognostic factors. *Ann Surg Oncol* 2010; **17**: 2494–502.
- 7 Fitzgerald RC, di Pietro M, Ragunath K, Ang Y, Kang J-Y, Watson P *et al.* British Society of Gastroenterology guidelines on the diagnosis and management of Barrett's oesophagus. *Gut* 2014; **63**: 7–42.
- 8 Weusten B, Bisschops R, Coron E, Dinis-Ribeiro M, Dumonceau J-M, Esteban J-M *et al.* Endoscopic management of Barrett's esophagus: European Society of Gastrointestinal Endoscopy (ESGE) Position Statement. *Endoscopy* 2017. doi:10.1055/s-0042-122140.
- 9 Shaheen NJ, Falk GW, Iyer PG, Gerson LB, American College of Gastroenterology. ACG Clinical Guideline: Diagnosis and Management of Barrett's Esophagus. *Am J Gastroenterol* 2016; **111**: 30–50.
- 10 Evans JA, Early DS, Fukami N, Ben-Menachem T, Chandrasekhara V, Chathadi K V. *et al.* The role of endoscopy in Barrett's esophagus and other premalignant conditions of the

- esophagus. *Gastrointest Endosc* 2012; **76**: 1087–1094.
- 11 Spechler SJ, Sharma P, Souza RF, Inadomi JM, Shaheen NJ. American Gastroenterological Association technical review on the management of Barrett's esophagus. *Gastroenterology* 2011; **140**: e18–e52.
 - 12 Kastelein F, van Olphen SH, Steyerberg EW, Spaander MCW, Bruno MJ, Biermann K *et al*. Impact of surveillance for Barrett's oesophagus on tumour stage and survival of patients with neoplastic progression. *Gut* 2015; **65**: 1–7.
 - 13 Verbeek RE, Leenders M, Ten Kate FJW, van Hillegersberg R, Vleggaar FP, van Baal JWPM *et al*. Surveillance of Barrett's Esophagus and Mortality from Esophageal Adenocarcinoma: A Population-Based Cohort Study. *Am J Gastroenterol* 2014; **109**: 1215–1222.
 - 14 Corley DA, Mehtani K, Quesenberry C, Zhao W, de Boer J, Weiss NS. Impact of endoscopic surveillance on mortality from Barrett's esophagus-associated esophageal adenocarcinomas. *Gastroenterology* 2013; **145**: 312–9.e1.
 - 15 Sturm MB, Wang TD. Emerging optical methods for surveillance of Barrett's oesophagus. *Gut* 2015; : 1–8.
 - 16 Brown H, Wyatt S, Croft S, Gale N, Turner A, Mulla A. Scoping the future: An evaluation of endoscopy capacity across the NHS in England. 2015.
 - 17 Chedgy FJQ, Subramaniam S, Kandiah K, Thayalasekaran S, Bhandari P. Acetic acid chromoendoscopy: Improving neoplasia detection in Barrett's esophagus. *World J Gastroenterol* 2016; **22**: 5753–5760.
 - 18 Thosani N, Abu Dayyeh BK, Sharma P, Aslanian HR, Enestvedt BK, Komanduri S *et al*. ASGE Technology Committee systematic review and meta-analysis assessing the ASGE Preservation and Incorporation of Valuable Endoscopic Innovations thresholds for adopting real-time imaging–assisted endoscopic targeted biopsy during endoscopic surveillance. *Gastrointest Endosc* 2016; **83**: 684–698.e7.
 - 19 Swager A, Curvers WL, Bergman JJ. Diagnosis by endoscopy and advanced imaging. *Best Pract Res Clin Gastroenterol* 2015; **29**: 97–111.
 - 20 NvisionVLE® Imaging System - NinePoint Medical.
<http://www.ninepointmedical.com/nvisionvle-imaging-system/> (accessed 1 Aug2017).
 - 21 Trindade AJ, Smith MS, Pleskow DK. The new kid on the block for advanced imaging in

- Barrett's esophagus: a review of volumetric laser endomicroscopy. *Therap Adv Gastroenterol* 2016; **9**: 408–16.
- 22 Saeian K, Staff DM, Vasilopoulos S, Townsend WF, Almagro U a, Komorowski R a *et al.* Unsedated transnasal endoscopy accurately detects Barrett's metaplasia and dysplasia. *Gastrointest Endosc* 2002; **56**: 472–8.
 - 23 Sugimoto H, Kawai T, Naito S, Yanagizawa K, Yamagishi T, Fukuzawa M *et al.* Surveillance of short-segment Barrett's esophagus using ultrathin transnasal endoscopy. *J Gastroenterol Hepatol* 2015; **30**: 41–45.
 - 24 Tanuma T, Morita Y, Doyama H. Current status of transnasal endoscopy using ultrathin videoscope for upper GI tract in the world. *Dig Endosc* 2016; **28**: n/a-n/a.
 - 25 Rodriguez SA, Banerjee S, Desilets D, Diehl DL, Farraye FA, Kaul V *et al.* Ultrathin endoscopes. *Gastrointest Endosc* 2010; **71**: 893–898.
 - 26 Sami SS, Dunagan KT, Johnson ML, Schleck CD, Shah ND, Zinsmeister AR *et al.* A randomized comparative effectiveness trial of novel endoscopic techniques and approaches for Barrett's esophagus screening in the community. *Am J Gastroenterol* 2015; **110**: 148–158.
 - 27 Moriarty JP, Shah ND, Rubenstein JH, Blevins CH, Johnson M, Katzka DA *et al.* Costs associated with Barrett ' s esophagus screening in the community : an economic analysis of a prospective randomized controlled trial of sedated versus hospital unsedated versus mobile community unsedated endoscopy. *Gastrointest Endosc* 2017. doi:10.1016/j.gie.2017.04.019.
 - 28 Iddan G, Meron G, Glukhovsky A, Swain P. Wireless capsule endoscopy. *Nature* 2000; **405**: 417–417.
 - 29 Fisher LR, Hasler WL. New vision in video capsule endoscopy: current status and future directions. *Nat Rev Gastroenterol Hepatol* 2012; **9**: 392–405.
 - 30 Wang A, Banerjee S, Barth BA, Bhat YM, Chauhan S, Gottlieb KT *et al.* Wireless capsule endoscopy. *Gastrointest Endosc* 2013; **78**: 805–815.
 - 31 Fernandez-Urien I, Carretero C, Armendariz R, Muñoz-Navas M. Esophageal capsule endoscopy. *World J Gastroenterol* 2008; **14**: 5254.
 - 32 Ciuti G, Menciassi A, Dario P. Capsule Endoscopy: From Current Achievements to Open Challenges. *IEEE Rev Biomed Eng* 2011; **4**: 59–72.
 - 33 Gora MJ, Sauk JS, Carruth RW, Lu W, Carlton DT, Soomro A *et al.* Imaging the upper

- gastrointestinal tract in unsedated patients using tethered capsule endomicroscopy. *Gastroenterology* 2013; **145**: 723–725.
- 34 Liao Z, Gao R, Xu C, Xu D-F, Li Z-S. Sleeve string capsule endoscopy for real-time viewing of the esophagus: a pilot study (with video). *Gastrointest Endosc* 2009; **70**: 201–209.
 - 35 Gora MJ, Sauk JS, Carruth RW, Gallagher K a, Suter MJ, Nishioka NS *et al*. Tethered capsule endomicroscopy enables less invasive imaging of gastrointestinal tract microstructure. *Nat Med* 2013; **19**: 238–40.
 - 36 Gupta N, Gaddam S, Wani SB, Bansal A, Rastogi A, Sharma P. Longer inspection time is associated with increased detection of high-grade dysplasia and esophageal adenocarcinoma in Barrett's esophagus. *Gastrointest Endosc* 2012; **76**: 531–8.
 - 37 Fuse® Full Spectrum Endoscopy®. <http://www.endochoice.com/Fuse> (accessed 1 Aug2017).
 - 38 Gralnek IM, Siersema PD, Halpern Z, Segol O, Melhem A, Suissa A *et al*. Standard forward-viewing colonoscopy versus full-spectrum endoscopy: An international, multicentre, randomised, tandem colonoscopy trial. *Lancet Oncol* 2014; **15**: 353–360.
 - 39 Hassan C, Senore C, Radaelli F, De Pretis G, Sassatelli R, Arrigoni A *et al*. Full-spectrum (FUSE) versus standard forward-viewing colonoscopy in an organised colorectal cancer screening programme. *Gut* 2017; **66**: 1949–1955.
 - 40 Clancy NT, Stoyanov D, James DRC, Di Marco A, Sauvage V, Clark J *et al*. Multispectral image alignment using a three channel endoscope in vivo during minimally invasive surgery. *Biomed Opt Express* 2012; **3**: 2567–78.
 - 41 Beg S, Wilson A, Ragunath K. The use of optical imaging techniques in the gastrointestinal tract. *Frontline Gastroenterol* 2016; **7**: 207–215.
 - 42 Longcroft-Wheaton G, Brown J, Basford P, Cowlshaw D, Higgins B, Bhandari P. Duration of acetowhitening as a novel objective tool for diagnosing high risk neoplasia in Barrett's esophagus: A prospective cohort trial. *Endoscopy* 2013; **45**: 426–432.
 - 43 Olliver JR, Wild CP, Sahay P, Dexter S, Hardie LJ. Chromoendoscopy with methylene blue and associated DNA damage in Barrett's oesophagus. *Lancet* 2003; **362**: 373–4.
 - 44 Ngamruengphong S, Sharma VK, Das A. Diagnostic yield of methylene blue chromoendoscopy for detecting specialized intestinal metaplasia and dysplasia in Barrett's esophagus: a meta-analysis. *Gastrointest Endosc* 2009; **69**: 1021–8.

- 45 Sharma P, Bergman JJGHM, Goda K, Kato M, Messmann H, Alsop BR *et al.* Development and Validation of a Classification System to Identify High-grade Dysplasia and Esophageal Adenocarcinoma in Barrett's Esophagus Using Narrow Band Imaging. *Gastroenterology* 2016; **150**: 591–598.
- 46 Kaneko K, Oono Y, Yano T, Ikematsu H, Odagaki T, Yoda Y *et al.* Effect of novel bright image enhanced endoscopy using blue laser imaging (BLI). *Endosc Int Open* 2014; **2**: E212–E219.
- 47 Osawa H, Yamamoto H, Miura Y, Sasao W, Ino Y, Satoh H *et al.* Blue laser imaging provides excellent endoscopic images of upper gastrointestinal lesions. *Video J Encycl GI Endosc* 2014; **1**: 607–610.
- 48 Miyake Y, Kouzu T, Takeuchi S, Yamataka S, Nakaguchi T, Tsumura N. Development of New Electronic Endoscopes Using the Spectral Images of an Internal Organ. In: *Proceedings of the IS&T/SID's Thirteen Color Imaging Conference*. Society for Imaging Science and Technology, 2005, pp 261–269.
- 49 Kodashima S, Fujishiro M. Novel image-enhanced endoscopy with i-scan technology. *World J Gastroenterol* 2010; **16**: 1043–1049.
- 50 Manfredi MA, Abu Dayyeh BK, Bhat YM, Chauhan SS, Gottlieb KT, Hwang JH *et al.* Electronic chromoendoscopy. *Gastrointest Endosc* 2015; **81**: 249–261.
- 51 Pohl J, May A, Rabenstein T, Pech O, Nguyen-Tat M, Fissler-Eckhoff A *et al.* Comparison of computed virtual chromoendoscopy and conventional chromoendoscopy with acetic acid for detection of neoplasia in Barrett's esophagus. *Endoscopy* 2007; **39**: 594–8.
- 52 Imagawa H, Oka S, Tanaka S, Noda I, Higashiyama M, Sanomura Y *et al.* Improved visibility of lesions of the small intestine via capsule endoscopy with computed virtual chromoendoscopy. *Gastrointest Endosc* 2011; **73**: 299–306.
- 53 Dung LR, Wu YY. A wireless narrowband imaging chip for capsule endoscope. *IEEE Trans Biomed Circuits Syst* 2010; **4**: 462–468.
- 54 von Holstein CS, Nilsson a M, Andersson-Engels S, Willen R, Walther B, Svanberg K. Detection of adenocarcinoma in Barrett's oesophagus by means of laser induced fluorescence. *Gut* 1996; **39**: 711–716.
- 55 Kara MA, Peters FP, Ten Kate FJW, Van Deventer SJ, Fockens P, Bergman JJGHM. Endoscopic video autofluorescence imaging may improve the detection of early neoplasia in

- patients with Barrett's esophagus. *Gastrointest Endosc* 2005; **61**: 679–685.
- 56 Curvers WL, Alvarez Herrero L, Wallace MB, Wong Kee Song L-M, Ragunath K, Wolfsen HC *et al.* Endoscopic tri-modal imaging is more effective than standard endoscopy in identifying early-stage neoplasia in Barrett's esophagus. *Gastroenterology* 2010; **139**: 1106–14.
 - 57 Curvers WL, van Vilsteren FG, Baak LC, Böhmer C, Mallant-Hent RC, Naber AH *et al.* Endoscopic trimodal imaging versus standard video endoscopy for detection of early Barrett's neoplasia: a multicenter, randomized, crossover study in general practice. *Gastrointest Endosc* 2011; **73**: 195–203.
 - 58 Xiong YQ, Ma SJ, Zhou JH, Zhong XS, Chen Q. A meta-analysis of confocal laser endomicroscopy for the detection of neoplasia in patients with Barrett's esophagus. *J Gastroenterol Hepatol* 2016; **31**: 1102–1110.
 - 59 Al-Rawhani MA, Beeley J, Cumming DRS. Wireless fluorescence capsule for endoscopy using single photon-based detection. *Sci Rep* 2016; **5**: 18591.
 - 60 Robles LY, Singh S, Fisichella PM. Emerging enhanced imaging technologies of the esophagus: spectroscopy, confocal laser endomicroscopy, and optical coherence tomography. *J Surg Res* 2015; **195**: 502–514.
 - 61 Lee JH, Wang TD. Molecular endoscopy for targeted imaging in the digestive tract. *Lancet Gastroenterol Hepatol* 2016; **1**: 147–155.
 - 62 Gora MJ, Suter MJ, Tearney GJ, Li X. Endoscopic optical coherence tomography: technologies and clinical applications [Invited]. *Biomed Opt Express* 2017; **8**: 2405.
 - 63 Trindade AJ, George BJ, Berkowitz J, Sejpal D V, McKinley MJ. Volumetric laser endomicroscopy can target neoplasia not detected by conventional endoscopic measures in long segment Barrett's esophagus. *Endosc Int open* 2016; **4**: E318-22.
 - 64 Swager A, Boerwinkel DF, de Bruin DM, Weusten BL, Faber DJ, Meijer SL *et al.* Volumetric laser endomicroscopy in Barrett's esophagus: A feasibility study on histological correlation. *Dis Esophagus* 2015; : 1–8.
 - 65 Suter MJ, Gora MJ, Lauwers GY, Arnason T, Sauk J, Gallagher K a. *et al.* Esophageal-guided biopsy with volumetric laser endomicroscopy and laser cautery marking: A pilot clinical study. *Gastrointest Endosc* 2014; **79**: 886–896.
 - 66 Ughi GJ, Gora MJ, Swager A, Soomro A, Grant C, Tiernan A *et al.* Automated segmentation

- and characterization of esophageal wall in vivo by tethered capsule optical coherence tomography endomicroscopy. 2016; **7**: 660–665.
- 67 Leggett CL, Gorospe EC, Chan DK, Muppa P, Owens V, Smyrk TC *et al.* Comparative diagnostic performance of volumetric laser endomicroscopy and confocal laser endomicroscopy in the detection of dysplasia associated with Barrett's esophagus. *Gastrointest Endosc* 2015; **83**: 880–888.e2.
 - 68 Lovat LB, Johnson K, Mackenzie GD, Clark BR, Novelli MR, Davies S *et al.* Elastic scattering spectroscopy accurately detects high grade dysplasia and cancer in Barrett's oesophagus. *Gut* 2006; **55**: 1078–83.
 - 69 Douplik A, Zanati S, Saiko G, Streutker C, Loshchenov M, Adler D *et al.* Diffuse reflectance spectroscopy in Barrett's Esophagus: Developing a large field-of-view screening method discriminating dysplasia from metaplasia. *J Biophotonics* 2014; **7**: 304–311.
 - 70 Perelman LT, Backman V. Light scattering spectroscopy of epithelial tissue: Principles and applications. In: *Handbook of Optical Biomedical Diagnostics*. SPIE PRESS, 2016.
 - 71 Wallace M, Perelman L, Backman V, Crawford J, Fitzmaurice M, Seiler M *et al.* Endoscopic detection of dysplasia in patients with Barrett's esophagus using light-scattering spectroscopy. *Gastroenterology* 2000; **119**: 677–682.
 - 72 Qiu L, Turzhitsky V, Chuttani R, Pleskow DK, Goldsmith JD, Guo L *et al.* Spectral Imaging With Scattered Light: From Early Cancer Detection to Cell Biology. *IEEE J Sel Top Quantum Electron* 2012; **18**: 1073–1083.
 - 73 Terry NG, Zhu Y, Rinehart MT, Brown WJ, Gebhart SC, Bright S *et al.* Detection of dysplasia in Barrett's esophagus with in vivo depth-resolved nuclear morphology measurements. *Gastroenterology* 2011; **140**: 42–50.
 - 74 Kim S, Heflin S, Kresty LA, Halling M, Perez LN, Ho D *et al.* Analyzing spatial correlations in tissue using angle-resolved low coherence interferometry measurements guided by co-located optical coherence tomography. *Biomed Opt Express* 2016; **7**: 1400.
 - 75 Qi J, Elson DS. A high definition Mueller polarimetric endoscope for tissue characterisation. *Sci Rep* 2016; **6**: 25953.
 - 76 Ba C, Palmiere M, Ritt J, Mertz J. Dual-modality endomicroscopy with co-registered fluorescence and phase contrast. *Biomed Opt Express* 2016; **7**: 3403.

- 77 Tan ACS, Tan GS, Denniston AK, Keane PA, Ang M, Milea D *et al.* An overview of the clinical applications of optical coherence tomography angiography. *Eye* 2017; : 1–25.
- 78 Kashani AH, Chen C-L, Gahm JK, Zheng F, Richter GM, Rosenfeld PJ *et al.* Optical coherence tomography angiography: A comprehensive review of current methods and clinical applications. *Prog Retin Eye Res* 2017; **60**: 66–100.
- 79 Wang L V, Yao J. A practical guide to photoacoustic tomography in the life sciences. *Nat Methods* 2016; **13**: 627–638.
- 80 Yang J-M, Li C, Chen R, Rao B, Yao J, Yeh C-H *et al.* Optical-resolution photoacoustic endomicroscopy in vivo. *Biomed Opt Express* 2015; **6**: 918.
- 81 Dong B, Chen S, Zhang Z, Sun C, Zhang HF. Photoacoustic probe using a microring resonator ultrasonic sensor for endoscopic applications. *Opt Lett* 2014; **39**: 4372–5.
- 82 Bai X, Gong X, Hau W, Lin R, Zheng J, Liu C *et al.* Intravascular optical-resolution photoacoustic tomography with a 1.1 mm diameter catheter. *PLoS One* 2014; **9**: e92463.
- 83 Yang J-M, Favazza CP, Yao J, Chen R, Zhou Q, Shung KK *et al.* Three-dimensional photoacoustic and ultrasonic endoscopic imaging of two rabbit esophagi. 2015; **9323**: 932334.
- 84 Zackrisson S, van de Ven SMWY, Gambhir SS. Light In and Sound Out: Emerging Translational Strategies for Photoacoustic Imaging. *Cancer Res* 2014; **74**: 979–1004.
- 85 Marcu L. Fluorescence Lifetime Techniques in Medical Applications. *Ann Biomed Eng* 2012; **40**: 304–331.
- 86 Pfefer TJ, Paithankar DY, Ponomarev JM, Schomacker KT, Nishioka NS. Temporally and spectrally resolved fluorescence spectroscopy for the detection of high grade dysplasia in Barrett's esophagus. *Lasers Surg Med* 2003; **32**: 10–16.
- 87 McGinty J, Galletly NP, Dunsby C, Munro I, Elson DS, Requejo-Isidro J *et al.* Wide-field fluorescence lifetime imaging of cancer. *Biomed Opt Express* 2010; **1**: 627–640.
- 88 Sun Y, Hatami N, Yee M, Phipps J, Elson DS, Gorin F *et al.* Fluorescence lifetime imaging microscopy for brain tumor image-guided surgery. *J Biomed Opt* 2010; **15**: 56022.
- 89 Cheng S, Rico-Jimenez JJ, Jabbour J, Malik B, Maitland KC, Wright J *et al.* Flexible endoscope for continuous in vivo multispectral fluorescence lifetime imaging. *Opt Lett* 2013; **38**: 1515–7.
- 90 Sparks H, Warren S, Guedes J, Yoshida N, Charn TC, Guerra N *et al.* A flexible wide-field

- FLIM endoscope utilising blue excitation light for label-free contrast of tissue. *J Biophotonics* 2015; **8**: 168–178.
- 91 Sun Y, Phipps JE, Meier J, Hatami N, Poirier B, Elson DS *et al*. Endoscopic fluorescence lifetime imaging for in vivo intraoperative diagnosis of oral carcinoma. *Microsc Microanal* 2013; **19**: 791–8.
 - 92 Chen J, Wong S, Nathanson MH, Jain D. Evaluation of Barrett esophagus by multiphoton microscopy. *Arch Pathol Lab Med* 2014; **138**: 204–12.
 - 93 Gu M, Kang H, Li X. Breaking the diffraction-limited resolution barrier in fiber-optical two-photon fluorescence endoscopy by an azimuthally-polarized beam. *Sci Rep* 2014; **4**: 3627.
 - 94 Jermyn M, Desroches J, Aubertin K, St-Arnaud K, Madore W-J, De Montigny E *et al*. A review of Raman spectroscopy advances with an emphasis on clinical translation challenges in oncology. *Phys Med Biol* 2016; **61**: R370–R400.
 - 95 Bergholt MS, Zheng W, Ho KY, Teh M, Yeoh KG, Yan So JB *et al*. Fiberoptic confocal raman spectroscopy for real-time in vivo diagnosis of dysplasia in Barrett's esophagus. *Gastroenterology* 2014; **146**: 27–32.
 - 96 Bergholt MS, Lin K, Zheng W, Huang Z, Lau D. In vivo, real-time, transnasal, image-guided {Raman} endoscopy: defining spectral properties in the nasopharynx and larynx. *J Biomed Opt* 2012; **17**: 77002.
 - 97 Wang Z, Liu Y, Gao L, Chen Y, Luo P, Wong KK *et al*. Use of multimode optical fibers for fiber-based coherent anti-Stokes Raman scattering microendoscopy imaging. *Opt Lett* 2011; **36**: 2967–2969.
 - 98 Légaré F, Evans CL, Ganikhanov F, Xie XS. Towards CARS Endoscopy. 2006; **14**: 4427–4432.
 - 99 Almond LM, Hutchings J, Lloyd G, Barr H, Shepherd N, Day J *et al*. Endoscopic Raman spectroscopy enables objective diagnosis of dysplasia in Barrett's esophagus. *Gastrointest Endosc* 2014; **79**: 37–45.
 - 100 Jermyn M, Mercier J, Aubertin K, Desroches J, Urmey K, Karamchandiani J *et al*. Highly accurate detection of cancer in situ with intraoperative, label-free, multimodal optical spectroscopy. *Cancer Res* 2017; **77**: 3942–3950.
 - 101 Lu G, Fei B. Medical hyperspectral imaging: a review. *J Biomed Opt* 2014; **19**: 10901.

- 102 Luthman AS, Dumitru S, Quiros-Gonzalez I, Joseph J, Bohndiek SE. Fluorescence hyperspectral imaging (fHSI) using a spectrally resolved detector array. *J Biophotonics* 2017; **10**: 840–853.
- 103 O'Connor JPB, Aboagye EO, Adams JE, Aerts HJWL, Barrington SF, Beer AJ *et al.* Imaging biomarker roadmap for cancer studies. *Nat Rev Clin Oncol* 2016; **14**: 169–186.
- 104 ICNIRP. ICNIRP guidelines on limits of exposure to incoherent visible and infrared radiation. *Health Phys* 2013; **71**: 804–819.
- 105 Coda S, Thompson AJ, Kennedy GT, Roche KL, Ayaru L, Bansi DS *et al.* Fluorescence lifetime spectroscopy of tissue autofluorescence in normal and diseased colon measured ex vivo using a fiber-optic probe. *Biomed Opt Express* 2014; **5**: 515–38.
- 106 Downs-Kelly E, Mendelin JE, Bennett AE, Castilla E, Henricks WH, Schoenfield L *et al.* Poor interobserver agreement in the distinction of high-grade dysplasia and adenocarcinoma in pretreatment Barrett's esophagus biopsies. *Am J Gastroenterol* 2008; **103**: 2333–40; quiz 2341.
- 107 Sharma P, Brill J, Canto M, DeMarco D, Fennerty B, Gupta N *et al.* White Paper AGA: Advanced Imaging in Barrett's Esophagus. *Clin Gastroenterol Hepatol* 2015; **13**: 2209–2218.
- 108 Atar M. Transnasal endoscopy: Technical considerations, advantages and limitations. *World J Gastrointest Endosc* 2014; **6**: 41.
- 109 Cellvizio: Our Flagship Product | Mauna Kea Technologies.
<http://www.maunakeatech.com/en/hospital-administrators/cellvizio-solution> (accessed 26 Apr2017).
- 110 PillCam ESO Capsule Endoscopy - Given Imaging. <http://www.givenimaging.com/en-int/Innovative-Solutions/Capsule-Endoscopy/Pillcam-ESO/Pages/default.aspx> (accessed 1 Aug2017).
- 111 Coletta M, Sami SS, Nachiappan A, Fraquelli M, Casazza G, Ragunath K. Acetic acid chromoendoscopy for the diagnosis of early neoplasia and specialized intestinal metaplasia in Barrett's esophagus: A meta-analysis. *Gastrointest Endosc* 2016; **83**: 57–67.
- 112 Mannath J, Subramanian V, Hawkey CJ, Ragunath K. Narrow band imaging for characterization of high grade dysplasia and specialized intestinal metaplasia in Barrett's esophagus: a meta-analysis. *Endoscopy* 2010; **42**: 351–9.

- 113 Song J, Zhang J, Wang J, Guo X, Yu S, Wang J *et al.* Meta-analysis of the effects of endoscopy with narrow band imaging in detecting dysplasia in Barrett's esophagus. *Dis Esophagus* 2015; **28**: 560–566.
- 114 Sharma P, Hawes RH, Bansal A, Gupta N, Curvers W, Rastogi A *et al.* Standard endoscopy with random biopsies versus narrow band imaging targeted biopsies in Barrett's oesophagus: a prospective, international, randomised controlled trial. *Gut* 2013; **62**: 15–21.
- 115 Maes S, Sharma P, Bisschops R. Review: Surveillance of patients with Barrett oesophagus. *Best Pract Res Clin Gastroenterol* 2016; **30**: 901–912.
- 116 Boerwinkel DF, Holz JA, Kara MA, Meijer SL, Wallace MB, Wong Kee Song L-M *et al.* Effects of autofluorescence imaging on detection and treatment of early neoplasia in patients with Barrett's esophagus. *Clin Gastroenterol Hepatol* 2014; **12**: 774–81.
- 117 Giacchino M, Bansal A, Kim RE, Singh V, Hall SB, Singh M *et al.* Clinical utility and interobserver agreement of autofluorescence imaging and magnification narrow-band imaging for the evaluation of Barrett's esophagus: a prospective tandem study. *Gastrointest Endosc* 2013; **77**: 711–8.